



Effects of Er:YAG and Er,Cr:YSGG laser irradiation and adhesive systems on microtensile bond strength of a self-adhering composite

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Abstract

This study was aimed to evaluate the effects of Er:YAG and Er,Cr:YSGG laser irradiation and adhesive systems on the microtensile bond strength of Fusio Liquid Dentin (FLD) which is a self-adhering composite (SAC). Twenty-four freshly extracted human molar teeth were collected, and the enamel was removed from the occlusal surface to obtain a flat dentin surface. Twenty-four teeth were randomly divided into eight groups: Group 1: only Fusio Liquid Dentin (FLD) (Petron Clinical, Orange, California, USA) was applied to the dentin surface; Group 2: 37% Phosphoricacid (i-GEL, Medicinos Linija UAB, Lithuania) + FLD; Group 3: Single Bond Universal (SBU) (3 M ESPE, Germany) + FLD; Group 4: Adper Easy One (AEO) (3 M ESPE, Germany) + FLD; Group 5: Er:YAG laser + AEO + FLD; Group 6: Er:YAG laser + SBU + FLD; Group 7: Er,Cr:YSGG laser + AEO + FLD; and Group 8: Er,Cr:YSGG laser + SBU + FLD. After thermocycling, $1 \times 1 \text{ mm}^2$ sticks were used for the μ TBS test ($n = 10$). Two sticks per group were used for SEM analysis. Fractured sample surfaces were evaluated using a stereomicroscope. Group 8 showed the highest μ TBS value (13.70 MPa), whereas Group 1 showed the lowest μ TBS value (5.60 MPa). There were no significant differences between Groups 2, 3, and 4 ($P = 0.324$), but Groups 5–8 showed statistically significant results that were higher than Groups 1–4 ($P = 0.012$). Adhesive failure mode was predominant followed by mixed failure. The evaluation of bonding of the FLD to dentin showed that the combined use of Er:YAG and Er,Cr:YSGG lasers with SBU and AEO on dentin surfaces improved the dentinal bond strength of the FLD.

Keywords Self-adhering composite · Composite · Surface preparation · Erbium laser · Microtensile bond strength

Introduction

Dental composites developed during the 1950s were mixtures of silicate glass particles acting as fillers within an acrylic monomer matrix polymerized during application. Some of those currently used are flowable, packable, microhybrid, with controlled shrinkage, smart, and nanohybrid composites, which were developed in the late 1990s and the early 2000s. These dental composites can be self-cured, UV light-cured, or visible light-cured. Flowable composites have the advantage of reaching inaccessible areas in Class II

cavities, which have a lower filler content than conventional composites [1–3].

Recent developments in adhesive and aesthetic dentistry have enabled the incorporation of adhesives into flowable resin composites. Self-adhering composites (SACs) have been introduced to overcome the complications of multiple-step procedures and to facilitate clinical placement. SACs do not require separate bonding and eliminate the need for adhesive application [4]. They are practical to use and can be quickly applied, making them advantageous in clinical practice. Accordingly, the use of self-adhering flowable resin composites during a single visit is preferable for uncooperative patients [5].

Dental composites require adhesive systems to bond with the hard dental tissues. There are two categories of adhesive systems, namely, self-etch (or etch-and-dry) and etch-and-rinse. Self-etch adhesive systems were preferred because of easier application and fewer application steps, resulting in reduced technique sensitivity [6, 7]. Etch-and-rinse adhesive systems are generally preferred for indirect restorations and

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when large areas of enamel are still present. Conversely, self-etch adhesives that are used on direct composite resin restorations supported by dentin provide more predictable bond strength and are recommended as a result [8].

Some factors, such as the type of adhesive system, restorative materials, and cavity preparation, affect the bond strength of resin to tooth structure. Recent research has emphasized the necessity of better tooth preparation to enhance the penetration of resin into the tooth structure. Dentin surface treatment using various agents, such as ultrasonic/sonic techniques, phosphoric acid, polyacrylic acid, and laser treatments, can affect the structural properties of dentin, which can change its microhardness, permeability, and solubility. The bonding strength between resin and dentin can be affected by these changes [9, 10].

Various types of lasers have been used for cutting hard dental tissue, for example, neodymium: yttrium aluminum garnet and the erbium group of lasers. Erbium-based lasers, Erbium; Yttrium Aluminum Garnet (Er:YAG) ($\lambda = 2.94 \mu\text{m}$) and Erbium, Chromium: Yttrium Scandium Gallium Garnet (Er,Cr:YSGG) ($\lambda = 2.78 \mu\text{m}$), known as hard tissue lasers, have high power and affect the surface morphology of dental tissues. Irregular surfaces opened dentinal tubules and prominent peritubular dentin without a smear layer were observed when the laser applied to dentin led to a microretentive surface, possibly favorable to bonding procedures [3, 11–15].

The Erbium lasers' handpiece did not touch the teeth during ablation. By eliminating vibrations this way, it protects the tooth from microscopic cracks and prevents the formation of grooves on the adjacent tooth surfaces [16]. The laser effects on dental tissues consist of thermomechanical wear and evaporation of water content. This causes expansion and disposal of organic and inorganic tissue contents and ultimately a surface with open dentinal tubules without a smear layer [17, 18]. The bond strength to tooth surfaces prepared with a laser is often confusing and accompanied by contradictory results. Some studies have shown that the bond strength to tooth surfaces prepared by low-power Er:YAG and Er,Cr:YSGG lasers is less than that to surfaces with acid-induced conditioning [17, 19].

In other studies, the laser efficiency in conditioning the tooth surface could become comparable to that with acid etching by changing some variables that belong to the laser equipment, such as its distance from the tooth surface and the output power [16]. Wajdowicz et al. suggested that laser irradiation was not effective in increasing bond strength, whereas Yazici et al. showed that the laser irradiation, as a tool for conditioning the dentin surfaces, increases the bond strength of SACs [20]. When the dental literature was examined, no study was found that investigated the effect of the combined use of erbium laser systems and different types of adhesives with SACs on the bond strength of dentin. Therefore, the primary objective of the present study was

to compare the bond strength of a SAC resin to the dentin surface after acid etching, adhesives, and laser conditioning. The null hypothesis was that there would be no significant differences between the bond strengths of the different surface treatments.

Materials and methods

After obtaining approval from Cumhuriyet University Clinical Research Ethics Committee (2017–05/01), 24 sound human third molars ($N = 24$) were collected, stored in saline solution at 4 °C, and were used within one month. All the root surfaces were cleaned to remove organic debris and deposits. Occlusal enamel was removed with a low-speed diamond saw (Isomet, Buehler, Lake Bluff, IL, USA). The sample surfaces were examined under a stereomicroscope to ensure that there was no enamel or pulp tissue on the surface. Standard dentin surfaces were obtained after sanding the occlusal surfaces on wet #180-grit SiC paper (60). #600-grit SiC was used to obtain a standardized smear layer.

Twenty-four teeth were randomly divided into eight groups, and three teeth were used in each group to obtain sticks for the μTBS test.

1. Group 1: Only Fusio Liquid Dentin (FLD, Pentron Clinical, Orange, CA, USA) was used as the SAC for this group. A stainless steel matrix was used to build a 3 mm height composite restoration on the dentin surface. The FLD was applied and cured with an LED curing device (Elipar S-10, 3 M ESPE, Germany) for 20 s.
2. Group 2: The flat dentin surfaces were etched with 37% phosphoric acid (i-GEL, Medicinos Linija UAB, Lithuania) for 15 s, rinsed, and then dried. FLD was applied and cured as described for Group 1.
3. Group 3: A universal adhesive Single Bond Universal (SBU, 3 M ESPE, Germany) was applied to the dentin surface for 20 s, gently dried for 5 s, and then light-cured for 10 s according to the manufacturer's instructions. Then, the FLD was applied and cured for 20 s as in Group 1.
4. Group 4: A self-etch adhesive Adper Easy One (AEO, 3 M ESPE) was applied to the dentin surface for 20 s, gently dried for 5 s, and then light-cured for 10 s. Following the adhesive application, FLD was applied and cured as in Group 1.
5. Group 5: Er:YAG laser (Smart 2940D Plus, Deka Laser, Florence, Italy) was applied to the dentin surface in non-contact mode with a tipless handpiece (Deka Laser, Florence, Italy). Laser energy was delivered at a wavelength of 2.94 μm , pulse duration of 150 μs for 30 s at 100 mJ energy output, 1 W, 10 Hz frequency, fluence 2.6 J/cm², and focal distance of 10 mm with a water

spray (70% water, 65% air), 5 mL/min with free-handed irradiation. Before and during sample irradiation, the output power of the laser beam was measured using a power meter (GS™ FieldMaster Power, Energy Analyzer, Coherent, Inc., Germany), and no power loss was observed. Then, AEO was applied to the dentin surface for 20 s, gently dried for 5 s, and then light-cured for 10 s. Following the adhesive application, FLD was applied and cured as in Group 1.

6. Group 6: After applying the Er:YAG laser according to the protocol of Group 5, SBU was applied to the dentin surface for 20 s, gently dried for 5 s, and then light-cured for 10 s. Then, FLD was applied and cured as in Group 1.
7. Group 7: Er,Cr:YSGG laser (Waterlase; Biolase Technology, San Clemente, CA, USA) was applied to the dentin surface. The laser parameters were as follows: power, 1.5 W; frequency, 30 Hz; cooling, air/water spray (70% water, 65% air), and water flow of nearly 14.5 mL/min; wavelength, 2780 nm; pulse duration, 60 μ s; fluence 4.5 J/cm²; and pulse energy 50 mJ in hard mode for 15 s [15]. The target was scanned homogeneously with a 600- μ m tip diameter from a distance of approximately 2 mm with free-handedness. Before and during sample irradiation, the output power of the laser beam was measured using a power meter (GS™ FieldMaster Power, Energy Analyzer, Coherent, Inc., Germany), and no power loss was observed. Then, AEO was applied to the dentin surface for 20 s, gently dried for 5 s, and then light-cured for 10 s. Following the adhesive application, FLD was applied and cured as in Group 1.
8. Group 8: After applying the Er,Cr:YSGG laser according to the protocol of Group 7, SBU was applied to the dentin surface for 20 s, gently dried for 5 s, and then light cured for 10 s. Following the adhesive application, FLD was applied and cured as in Group 1.

Aging procedure

All specimens in the groups were thermocycled between 5 °C and 55 °C (5000 cycles; Gokceler Mechanical, Sivas, Turkey) with a dwell time of 30 s in a water bath. The transfer time was 5 s to see aging affects on μ TBSs.

Microtensile bond strength (μ TBS) testing

A nontrimming technique was used for microtensile testing. The specimens were sectioned (in the x and y directions) using a diamond saw. The sticks obtained from three teeth were measured using a digital caliper, and the closest 10 sticks (1 \times 1 mm²) were selected for each group ($n=10$). Samples were stored in distilled water for 24 h at room temperature under daylight, and then attached to a universal testing machine (LF Plus, LLOYD Instruments,

Ametek Inc., England) with cyanoacrylate adhesive and an accelerator (404 Super cyanoplast, 404 Kimya Sanayi, İstanbul, Türkiye). The samples were tested under tension at a crosshead speed of 0.5 mm/min until failure. The μ TBS values were then calculated and expressed in MPa.

Fractured sample surfaces were evaluated using a stereomicroscope (Olympus SZX7, Germany) to examine failure types such as adhesive, mix, or cohesive failure.

For each group, two sticks were randomly allocated for examination using a scanning electron microscope (SEM).

Statistical analysis

The obtained data were analyzed using the IBM SPSS Statistics 25.0 (IBM Corporation) software. Analysis of microtensile bond strength was performed using one-way ANOVA, and multiple comparisons were performed using Tukey's posthoc test. Statistical significance was set at $P < 0.05$.

Results

The mean bond strength values of the groups are shown in Table 1. Group 8 showed the highest μ TBS value (13.70 MPa), whereas Group 1 showed the lowest μ TBS value (5.60 MPa). There were no significant differences between Groups 6 and 8, and both groups showed statistically significant differences compared with Groups 5 and 7 ($P=0.001$). Group 7 showed higher values than Group 5, but the difference was not statistically significant. There were no significant differences between Groups 2, 3, and 4 ($P=0.324$), but Groups 5 and 7 showed significantly higher results than these groups ($P=0.001$). Group 2 showed higher results than Groups 3 and 4, but there were no statistically significant differences. Other groups (2–8) showed higher results than Group 1, and they were statistically different ($P=0.001$).

In SEM analysis, it was hard to see hybrid layer and resin tags especially in Group 1 (Fig. 1 (1a, 1b)). As there were fully bonded areas there were cracks in most samples (Fig. 1 (2a, 2b)). The FLD resin showed limited penetration to tubules, and therefore, shortened resin tags were seen (Fig. 2). There was discontinuity of the hybrid layer with broken resin tags in Groups 3 and 4 (Fig. 1 (3a, 3b)). Adhesive failure mode was predominant (between adhesive and dentin), followed by mixed failure in all groups (Table 2). Example SEM images of fracture types are shown in Fig. 3.

Table 1 The mean, standard deviations, maximum and minimum values of the groups. The letters show the statistically differences between groups

Groups	n	Mean \pm Std. dev. (MPa)	Min. (MPa)	Max. (MPa)
Group 1 (FLD)	10	5.60 ^A \pm 1.34	3.93	7.68
Group 2 (Acid-etch + FLD)	10	8.91 ^B \pm 1.23	6.74	10.67
Group 3 (SBU + FLD)	10	7.81 ^B \pm 0.69	6.74	8.91
Group 4 (AEO + FLD)	10	7.64 ^B \pm 0.64	6.27	8.42
Group 5 (Er: YAG + AEO + FLD)	10	11.26 ^C \pm 1.62	9.02	14.42
Group 6 (Er: YAG + SBU + FLD)	10	13.64 ^{C,D} \pm 1.34	11.58	15.40
Group 7 (Er:Cr:YSGG + AEO + FLD)	10	11.44 ^C \pm 1.58	9.32	13.67
Group 8 (Er:Cr:YSGG + SBU + FLD)	10	13.70 ^D \pm 1.91	11.31	16.60

Std. dev. standard deviation, *Min* minimum, *Max* maximum

*In each column, groups with different capital superscripts are significantly different ($P < 0.05$)

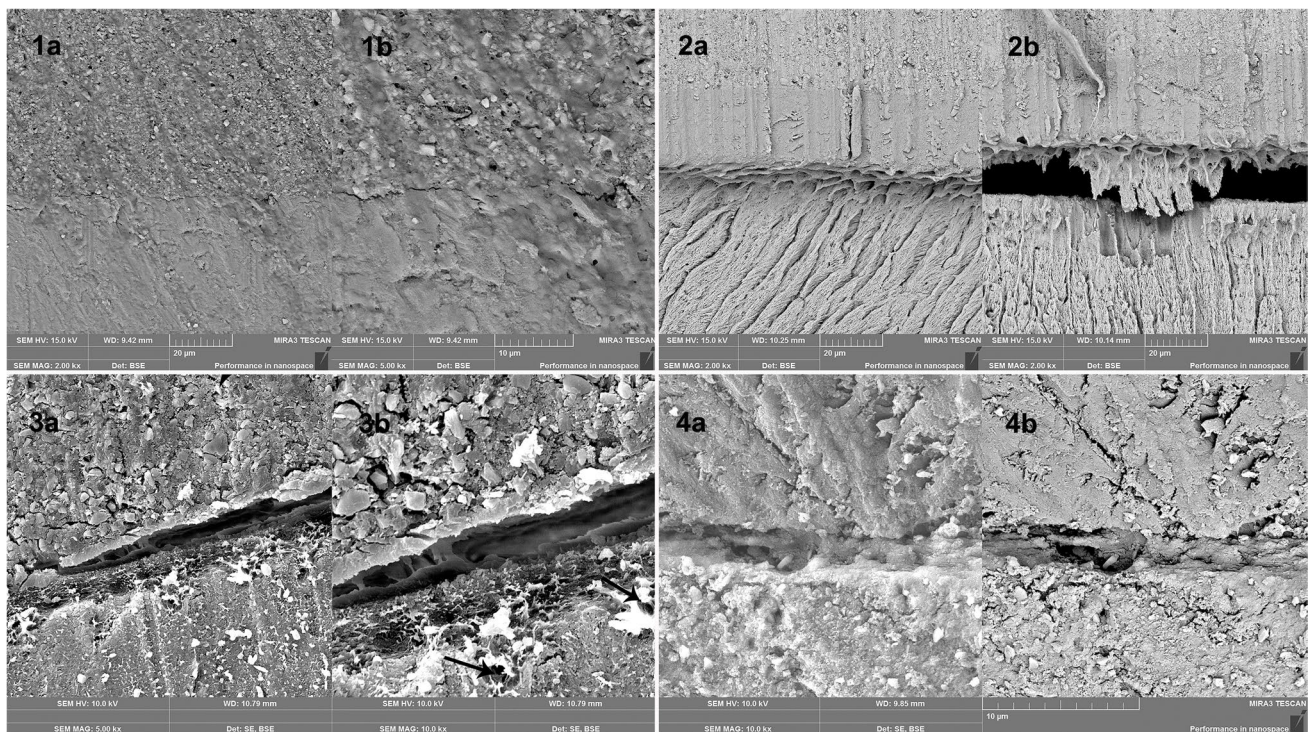


Fig. 1 (1a, 1b) Hybrid layer and resin tags could not be seen easily in Group 1. (2a, 2b) There were fully bonded areas but most of them were cracked in Group 2. (3a, 3b) There were opened dentin tubules

(black arrow) in lased surfaces but unable to detect full-filled resin tags in Group 5. (4a, 4b) Discontinuity of a hybrid layer with broken resin tags in Group 3

Discussion

Bonding to dental tissues is considered to be of great importance in restorative dentistry to reestablish aesthetics and function within the concepts of maximum preservation of the tooth structure. Different dentin surface treatments can influence the effectiveness of dentin bonding systems [21, 22].

Lasers have been recommended to increase the adhesion of the resin to tooth structures [20]. It has also been used to etch or modify the surface of teeth as a substitute

for acid etching. However, the effectiveness of this technique is controversial. While some researchers support the preparation or etching ability of laser to dentin [23–25], others deny its efficacy [26–28].

When the electromicrographs (SEM) were evaluated, the morphological features of the conventionally prepared dentin surface (i.e., diamond bur) differed from those of dentin treated with erbium lasers [29]. In the former, there was an evident smear layer obliterating the dentinal tubule entrances, whereas in the latter, the laser-irradiated dentin presented an irregular surface without a smear layer, with open dentinal tubules and prominent peritubular dentin,

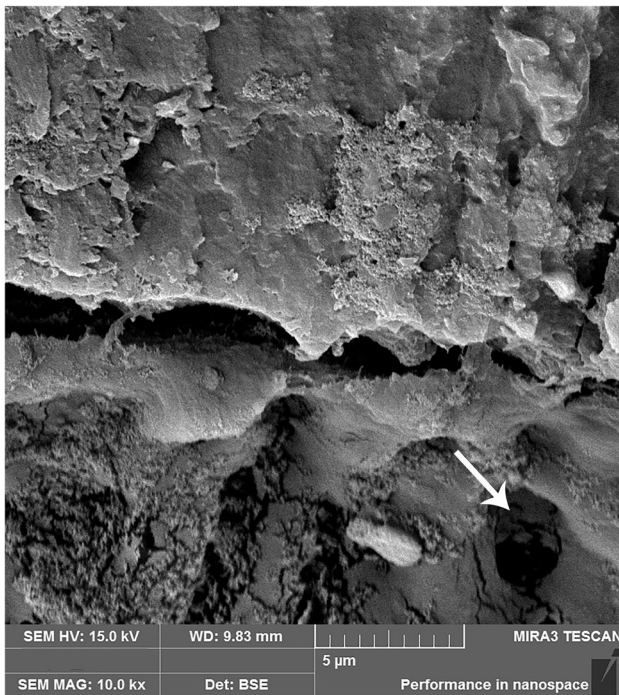


Fig. 2 A cracked resin-dentin interface with a thin hybrid layer and shortened resin tags (white arrow) in acid-etched and laser surface in Group 8

Table 2 Failure distribution of groups

Groups	Adhesive failure (%)	Cohesive failure (%)	Mix failure (%)
Group 1 (FLD)	90	-	10
Group 2 (Acid-etch + FLD)	90	-	10
Group 3 (SBU + FLD)	70	-	30
Group 4 (AEO + FLD)	60	-	40
Group 5 (Er:YAG + AEO + FLD)	50	10	40
Group 6 (Er:YAG + SBU + FLD)	60	20	20
Group 7 (Er,Cr:YSGG + AEO + FLD)	70	-	30
Group 8 (Er,Cr:YSGG + SBU + FLD)	60	-	40

suggesting a microretentive morphology favorable to bonding procedures [30–34].

This study determined how Er:YAG and Er,Cr:YSGG lasers, which are used for ablation of mineralized tissue in dentistry, affect the bonding of composites on the irradiated dentin surface. The irradiation parameters proposed were lower than those reported in previous studies because the goal was to modify the eroded dentin surface without necessarily promoting ablation or cavity formation [12, 35].

Another factor affecting the success of composite restorations is the use of complicated adhesive systems to treat

dentin, which requires various steps and therefore is associated with technique sensitivity, leading to increased chair time. By eliminating etching, rinsing, priming, and bonding [36], new self-adhesive flowable resins may help to reduce chair time and can be useful in uncooperative patient treatment, such as in children. [5, 20].

Self-adhesive flowable composites differ in their composition and feature various functional monomers. Fusio Liquid Dentin chemically bonds to hydroxyapatite using 4-methacryloxyethyl trimellitic acid (4-META), which can partially demineralize dentin and form ionic bonds between its carboxylate groups and calcium in hydroxyapatite [37].

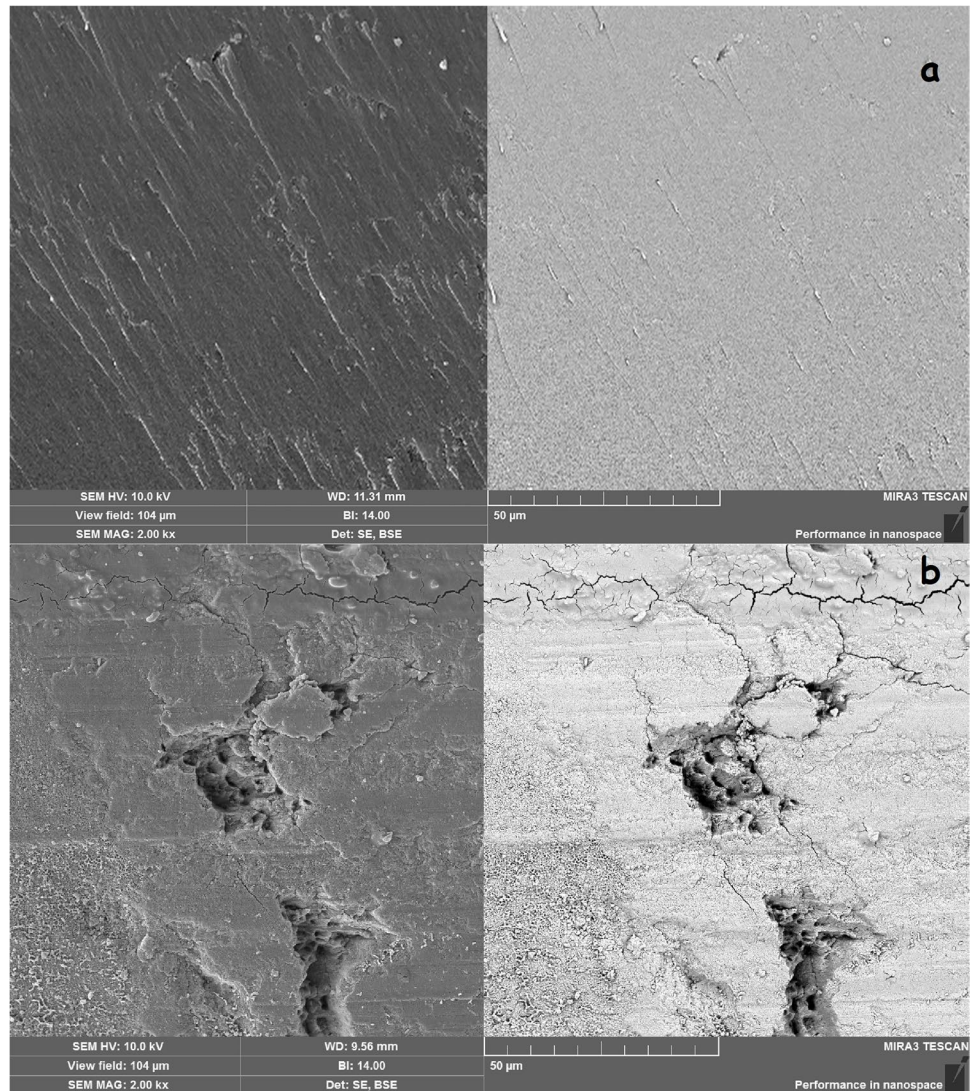
In the present study, Group 1 (FLD) showed the lowest μ TBS. FLD partially demineralizes dentin and obtains the remaining hydroxyapatite partially attached to collagen within a submicron hybrid layer. It can be concluded from research that being less acidic causes decreased bond strength [2, 38, 39]. In previous studies, FLD showed decreased bond strengths like other SACs and the authors of this paper are of the same opinion that FLD showed statistically less μ TBS values than that of the other groups [2, 7, 38–41]. A possible reason for the low bond strength is the less acidic effect and the occurrence of water droplets between the resin and dentin [42]. In addition, the blockage of dentin tubules with the smear layer that occurred after SiC preparation may account for the improper penetration of the FLD. When FLD was applied to dentin, thin and sparse tags were seen in some studies [7, 42], and the hybrid layer and resin tags could not be viewed [38, 42].

According to the SEM analysis, in Group 1, a thin hybrid layer that was difficult to observe was present, which showed discontinuity in most parts of the resin-dentin bonding surface. (Fig. 1) In Groups 2–8, a short hybrid layer can be observed with short resin tags. (Fig. 2) Adhesive failure mode was predominant, followed by mixed failure. Adhesive failures were observed in groups that showed decreased μ TBS values with weak micromechanical bonding to dentin, and mixed fractures were mostly observed in groups that showed increased μ TBS values (Table 2) (Fig. 3).

Different surface treatments have been widely used to obtain increased μ TBS values, such as surface acid etching, laser irradiation, sandblasting, abrasion with a diamond bur followed by silica coating, and using intermediate bonding agents [43, 44].

Acid etching is used for demineralization of subsurface intact dentin and complete removal of the smear layer [39]. Acid etching not only exposes the network of the collagen matrix in dentin but also makes the dentinal tubules wide and open [40]. When acid is applied to the surface, it removes the smear layer and partially demineralizes the dentin to allow adhesive penetration into the collagen fibrils [15]. Shafiei and Saadat [6] evaluated the micromorphology and shear bond strength of SAC and found that the

Fig. 3 SEM images of failure types. (a) Adhesive and (b) mix failures



phosphoric acid-treated group had the highest bond strength. Altunsoy et al. [40] evaluated the effect of different surface treatments on the μ TBS of two different SACs to dentin and showed that acid etching increased the μ TBS of SACs. Poitevin et al. [41] evaluated the bonding effectiveness of SAC and reported that phosphoric acid etching of dentin/enamel significantly improved the bonding effectiveness of SAC. In Group 2, phosphoric acid (37%) was used, and increased values were observed in the μ TBS values of SAC. The current values are in line with the studies that acid etching before applying SAC can increase the bond strength of these materials.

The bond strength between the dental substrate and adhesive system is one of the most important factors to consider for successful restorative treatment. When an adhesive agent was used with SACs as in conventional composites, increased μ TBS values were observed in both Groups 3 and 4. A universal bond (SBU) was used in Group 3, and

a self-etch bond (AEO) was used in Group 4. Cengiz and Unal [38] compared two SACs with universal adhesives in two modes, total-etch and self-etch, and Single Bond Universal showed higher μ TBS values than FLD when FLD was applied alone in their study. Sismanoglu [44] tested an SAC for the repair of composites and showed increased μ TBS values in the use of Single Bond Universal with SAC.

Studies on SACs have reported that the bond strengths of SACs are improved with the use of an adhesive layer [1, 5, 36, 45]. Although SBU showed higher μ TBS values than AEO, there were no significant differences between them. (Table 1) Two adhesives, SBU and AEO, include Vitrebond copolymer (1–5%), which may also provide chemical bonding to hydroxyapatite. For self-etch adhesives, chemical bonding between polycarboxylic monomers and hydroxyapatite plays a crucial role in their bonding mechanism [46]. Depending upon the acid dissociation constants (pKa values), the etching aggressiveness of self-etch adhesive

systems can also be classified into “strong” ($\text{pH} < 1$), “intermediately strong” ($\text{pH} \approx 1.5$), “mild” ($\text{pH} \approx 2$), and “ultramild” ($\text{pH} \geq 2.5$). The bonding used in this study SBU had a pH of 2.7 and AEO had a pH 0.8–1. As expected, the strong acidity of the AEO could solve more smears and provide increased bond strength. However, no statistically significant differences were observed between the two bonding agents. AEO contains a methacrylohexyl phosphate monomer, and SBU contains a methacryloyloxi-decyl-dihydrogen-phosphate (MDP) monomer. MDP has a phosphoric-acid functional group that interacts chemically with hydroxyapatite crystals and forms stable calcium phosphate and calcium carboxylate salts, with a limited surface-decalcification effect. MDP has another methacrylate polymerizable group that is responsible for the curing potential and a 10-carbon chain group that separates the other active groups. The carbon spacer influences the hydrophobicity-hydrophilicity balance, monomer flexibility, solubility, and wetting. One of the most important reasons for bond durability is the interaction between MDP and the additional chemical interaction of dentin [47]. This seems to be the most likely cause of the nearest μTBS values of SBU and AEO, and the authors of this study are in the same opinion as Pashaev et al. [46] who reported that when universal adhesives were used in the self-etch application mode, SBU exhibited μTBS values similar to AEO.

In the current study, all laser-treated groups (Groups 5–8) showed significantly higher bonding strength values than the untreated groups (Groups 1–4). This is because of three features of the tooth surface as a result of laser irradiation: (1) roughness of the surface, (2) open dentinal tubules, and (3) the lack of a smear layer on the surface [48]. Er:YAG and Er,Cr:YSGG lasers showed similar bond strength values when used with the same adhesive agent. There were no statistically significant differences between Groups 5 and 7 and between Groups 6 and 8 in terms of bond strength. (Table 1). However, the groups in which SBU was applied together with laser applications (Groups 6 and 8) showed higher bond strength values than the groups in which AEO was applied with laser applications (Groups 5 and 7). While Group 8 differed significantly from Group 7, there was no statistically significant difference between Groups 6 and 5.

Altunsoy et al. [40] evaluated the influence of various surface preparations on the μTBS of two SACs and reported that Er:YAG laser irradiation increased μTBS when using FLD. Nahas et al. [14] investigated the bond strength of SAC in Er:YAG laser-pretreated surfaces and reported that with an optimal energy level, the Er:YAG laser may enhance the bond strength of SAC. Yazici et al. [20] evaluated the effect of Er:YAG laser preparation on the μTBS of SAC and found increased bond strengths in their study. Zarabian et al. [49] reported increased bond strength after evaluating the bond strength of an SAC to Er,Cr: YSGG-treated enamel. Moslemi et al. [13]

evaluated the effect of an Er, Cr: YSGG laser on the bond strength of SAC and reported that laser irradiation can improve bond strength. The authors of the present study are in the same opinion as in the aforementioned literature that laser irradiation can improve bond strengths of SACs [13, 14, 20, 40]. Memarpour et al. [1] evaluated the effect of laser preparation on the adhesion of SAC and found lower bond strength values. The main reason for the decreased result may be the SACs monomers used in that study, and the laser parameters may have an effect on the bond strength. In the current study, the application of dental adhesives to dentin surfaces after laser application increased the bond strength of SAC. This increase may have been due to the content of Vitrebond copolymer (1%–5%) (which might also provide chemical bonding to hydroxyapatite) and the adhesives that have acidic pH values (SBU pH 2.7 and AEO pH 0.8–1).

This study has some limitations. Firstly, this study conducted an in vitro study design. Thus, it is not possible to fully reflect the oral environment. Another limitation is that different parameters have been tested in the current study as it may be possible to argue that it would effect the reliability of the findings. However, it should be noted that several methodologies have been performed in previous studies and all tested parameters in the current study have also been confirmed in previous studies. Therefore, further in vitro and in vivo studies with eroded dentin are necessary to assess the new parameters of Er:YAG and Er,Cr:YSGG laser irradiation. In addition, Erbium laser application was combined with adhesive systems in this study. There is a need for studies where laser systems are applied only with SACs.

Conclusion

According to the results of the current in vitro study, it is clear that FLD provides insufficient adhesion when used alone. Therefore, an alternative surface preparation is advised to have better adhesion. When the bonding of the FLD to dentin was evaluated, the combined use of Er:YAG and Er,Cr:YSGG lasers with SBU and AEO on dentin surfaces improved the dentinal bond strength of the FLD.

Author contribution Alper Kaptan contributed to conception, design, collecting data, statistical analysis, and writing some parts of manuscript. Fatih Oznurhan contributed to conception, design, data acquisition and interpretation, and SEM analysis writing of the manuscript.

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Data availability The data that support the findings of this study are available from the corresponding author [FO], upon reasonable request.

Declarations

Ethics approval This study was approved by Cumhuriyet University Clinical Research Ethic Committee (2017–05/01).

Conflict of interest The authors declare no competing interests.

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