BIOCHEMISTRY AND MOLECULAR BIOLOGY IN THE POST GENOMIC ERA

METABOLOMICS And CLINICAL APPROACH

SEVGI DURNA DAŞTAN TANER DAŞTAN editors

Ť

Biochemistry and Molecular Biology in the Post Genomic Era



No part of this digital document may be reproduced, stored in a retrieval system or transmitted in any form or by any means. The publisher has taken reasonable care in the preparation of this digital document, but makes no expressed or implied warranty of any kind and assumes no responsibility for any errors or omissions. No liability is assumed for incidental or consequential damages in connection with or arising out of information contained herein. This digital document is sold with the clear understanding that the publisher is not engaged in rendering legal, medical or any other professional services.

Biochemistry and Molecular Biology in the Post Genomic Era

Structure and Function of Cytochromes

Elaine J. Mathis (Editor) 2022. ISBN: 979-8-88697-102-6 (Hardcover) 2022. ISBN: 979-8-88697-122-4 (eBook)

Laboratory Guide: Concepts and Protocols for Practical Courses in Biochemistry and Molecular Biology

Shafat Ahmad Latoo, M.S., PhD (Editor) Mohammad Asif Shah, M.S., PhD (Editor) 2022. ISBN: 978-1-68507-862-1 (Hardcover) 2022. ISBN: 979-8-88697-028-9 (eBook)

Omics for Animal Sciences: Principles and Approaches

Asif Nadeem (Author) Maryam Javed (Author) 2022. ISBN: 978-1-68507-722-8 (Hardcover) 2022. ISBN: 978-1-68507-901-7 (eBook)

The Future of Metabolic Engineering

Abhishek Sharma, PhD (Editor) Dhruti Amin, PhD (Editor) 2022. ISBN: 978-1-68507-362-6 (Hardcover) 2022. ISBN: 978-1-68507-464-7 (eBook)

Recent Advances in Computer Aided Drug Designing

Akhil Varshney, PhD, Ashutosh Mani, PhD (Editors) 2021. ISBN: 978-1-53619-739-6 (Hardcover) 2021. ISBN: 978-1-53619-904-8 (eBook)

More information about this series can be found at https://novapublishers.com/product-category/series/biochemistry-and-molecularbiology-in-the-post-genomic-era/

Sevgi Durna Daştan and Taner Daştan Editors

Metabolomics and Clinical Approach



Copyright © 2023 by Nova Science Publishers, Inc.

DOI: https://doi.org/10.52305/FZMO0355

All rights reserved. No part of this book may be reproduced, stored in a retrieval system or transmitted in any form or by any means: electronic, electrostatic, magnetic, tape, mechanical photocopying, recording or otherwise without the written permission of the Publisher.

We have partnered with Copyright Clearance Center to make it easy for you to obtain permissions to reuse content from this publication. Please visit copyright.com and search by Title, ISBN, or ISSN.

For further questions about using the service on copyright.com, please contact:

Phone: +1-(978) 750-8400

Copyright Clearance Center Fax: +1-(978) 750-4470

E-mail: info@copyright.com

NOTICE TO THE READER

The Publisher has taken reasonable care in the preparation of this book but makes no expressed or implied warranty of any kind and assumes no responsibility for any errors or omissions. No liability is assumed for incidental or consequential damages in connection with or arising out of information contained in this book. The Publisher shall not be liable for any special, consequential, or exemplary damages resulting, in whole or in part, from the readers' use of, or reliance upon, this material. Any parts of this book based on government reports are so indicated and copyright is claimed for those parts to the extent applicable to compilations of such works.

Independent verification should be sought for any data, advice or recommendations contained in this book. In addition, no responsibility is assumed by the Publisher for any injury and/or damage to persons or property arising from any methods, products, instructions, ideas or otherwise contained in this publication.

This publication is designed to provide accurate and authoritative information with regards to the subject matter covered herein. It is sold with the clear understanding that the Publisher is not engaged in rendering legal or any other professional services. If legal or any other expert assistance is required, the services of a competent person should be sought. FROM A DECLARATION OF PARTICIPANTS JOINTLY ADOPTED BY A COMMITTEE OF THE AMERICAN BAR ASSOCIATION AND A COMMITTEE OF PUBLISHERS.

Library of Congress Cataloging-in-Publication Data

Names: Durna Daştan, Sevgi, editor. | Daştan, Taner, editor. Title: Metabolomics and clinical approach / Sevgi Durna Daştan, editor, Associate Professor, Department of Biology, Division of Molecular Biology, Faculty of Science, Sivas Cumhuriyet University, Sivas, Turkey, Taner Daştan, PhD, editor, Faculty of Science and Department of Biochemistry, Sivas Cumhuriyet University, Sivas, Turkey. Description: New York : Nova Science Publishers, [2023] | Series: Biochemistry and molecular biology in the post genomic era | Includes bibliographical references and index. | Identifiers: LCCN 2023041393 (print) | LCCN 2023041394 (ebook) | ISBN 9798891130951 (hardcover) | ISBN 9798891131811 (adobe pdf) Subjects: LCSH: Metabolites. Classification: LCC QP171 .M48223 2023 (print) | LCC QP171 (ebook) | DDC 612.3/9-.dc23/eng/20231010 LC record available at https://lccn.loc.gov/2023041393

LC ebook record available at https://lccn.loc.gov/2023041394

Published by Nova Science Publishers, Inc. † New York

Contents

Preface	ix
Chapter 1	The Concept of Metabolomics1 Sinan Vıcıl and Tünay Karan
Chapter 2	Metabolomics and BMI21 Bade Ertürk Arik
Chapter 3	Metabolites Associated with Age and Aging31 Ali Sağlik
Chapter 4	Geno-Metabolomics: Genotype Effects on Metabolites47 Şeyda Kaya
Chapter 5	Pharmaco-Metabolomics: Drug Effects on Metabolites55 Eda Sonmez Gurer
Chapter 6	The Use of HPLC in Metabolomics Research65 Tunay Karan and Sinan Vicil
Chapter 7	The Scope of LC-MS-Based Metabolomics83 Pelin Koparir and İnanç Baral
Chapter 8	Databases for Metabolomics Research91 Dilek Arslan Ateşşahin and Yusuf Muhammed Durna
Chapter 9	Metabolomic Research: Past, Present, and Future103 Gonca Gulbay
Chapter 10	Metabolomics in Traumatic Brain Injuries115 Mustafa Karademir

Contents

Chapter 11	The Clinical Impact and Use of Metabolomics in Diabetes Research123 Melike Mercan Başpinar
Chapter 12	The Current State and Future Perspectives of Metabolomics in Orthopedics135 Anıl Aydin
Chapter 13	Metabolomics and Metabolic Profiling in Embryos for Viability Assessment145 Savaş Karakuş
Chapter 14	Metabolomics in the Diagnosis of Pregnancy Complications 155 Nazan Yurtcu and Dilay Karademir
Chapter 15	Metabolomic Markers and Their Clinical Uses in Neurology169 Gülay Soykök
Chapter 16	Metabolomics in Immunology Research185 Semih Dalkiliç and Lütfiye Kadioğlu Dalkiliç
Chapter 17	The Clinical Uses of Mass Spectrometry-Based Approaches in Neurology and Psychiatry197 Suna Dagli Atalay
Chapter 18	Nutritional Metabolomics and Their Implications in Clinical Studies215 Semra Türkoğlu
Chapter 19	Metabolome in Urology Practices: Signatures in Plasma and Urine225 Aydemir Asdemir
Chapter 20	Metabolomic Applications in Food Science and Nutrition241 Ayse Burcu Aktas
Chapter 21	The Use of Metabolomics in Neonatology257 Murat Doğan
Chapter 22	Metabolomic Profiles of Body Fluids in Preterm Births271 Seda Yilmaz Semerci and Adviye Cakil Saglik

Chapter 23	The Use of Metabolomics in Diagnosing and Evaluating Inborn Metabolism Errors Adviye Cakil Saglik	287
Chapter 24	Metabolomics in Biomarker Identification for Cardiovascular Diseases Mesude Avci	297
Index		311

Chapter 20

Metabolomic Applications in Food Science and Nutrition

Ayse Burcu Aktas*, PhD

Department of Biochemistry, Faculty of Science, Sivas Cumhuriyet University, Sivas, Turkey

Abstract

The definition of metabolomics encloses an efficient description of small molecule metabolites present in different biological matrices. Metabolomics has shown up as a prominent tool for various areas including health diseases, drug research, plant science, food science, and nutrition. The metabolomics has recently been used to investigate some parameters in particular for food quality, processing, and the safety of both raw materials and final products. Food metabolomics has also a position in human nutrition research. This book chapter focuses on the novel and potential applications of metabolomics in food science and nutrition by summarizing some metabolomic analysis of food components, metabolomics from the preferential, presumable, and informative approaches in food processing, food microbiology, food safety, food quality, and benefits of foodomics in human nutrition and related health diseases.

Keywords: metabolomics, foodomics, food science, food processing, food analyses, nutrition

^{*} Corresponding Author's Email: burcuaktas@cumhuriyet.edu.tr.

In: Metabolomics and Clinical Approach Editors: Sevgi Durna Daştan and Taner Daştan ISBN: 979-8-89113-095-1 © 2023 Nova Science Publishers, Inc.

Introduction

Metabolomics is a discipline to explain the performances of metabolites by identifying, quantifying, and detailing their mechanisms of action. There are several advanced instruments and techniques to investigate the metabolomes of various samples, including fundamentally nuclear magnetic resonance (NMR), mass spectrometry (MS), and vibrational spectrometry (VS) or combinations of other techniques. Generally, all these instrumental techniques are aimed to isolate, determine, quantify, characterize structures, and explain their functions on the metabolic pathways. Since metabolomes are complex molecules, it is highly needed to conduct many of these techniques together to better understand the whole mechanism (Fraga-Corral et al., 2022; Zhang et al., 2012).

Metabolomics has great potential to figure out the major problems of food science. Moreover, metabolomics is considered as an effective technique for further agricultural needs (Green et al., 2007; Hall et al., 2008). The engagements of informative, discriminative, and predictive metabolomics have been used for food quality, process, nutrition, and food component analysis (Cevallos-Cevallos et al., 2009). This book chapter covers a review of current metabolomics applications in food quality, safety, microbiology, processing, and nutrition.

Metabolomics in Food Science

Analysis of Food Metabolomics

The analysis of food metabolomics has been used in food safety control, including foodborne pathogens, toxins, some allergens, pesticides, antibiotics, food quality (in terms of organoleptic properties and nutritional value), food authenticity, and food traceability (Cook and Nightingale, 2018).

There are six main steps for metabolomics analysis including sample preparation, metabolite extraction, derivatization, metabolite separation, detection, and data treatment. In general, the essential steps for metabolomics analysis are detection and data analysis. The steps could be selected according to the purpose of the study, sample type, instrument type, and detection method (Pinu, 2015). Some of the metabolomics studies related to different food samples are summarized in Table 1. Since the metabolomic analyses of foods are quite varied, most studies could be classified as discriminative. The

informative metabolomic analyses are mostly interested in the recognition and quantification of certain metabolites to obtain detailed information about the food sample. The untargeted metabolomics solely focuses on the identification of some groups of metabolites to obtain patterns or fingerprints, whereas targeted metabolomics demands higher purification and selective extraction stages (Alonso et al., 2015). Some metabolomics studies are predictive means that statistical models are constructed to estimate a variable that is difficult to quantify based on the metabolite profile and abundance (Table 1).

Sample and purpose	Analysis Type	Instrumentation	Data Analysis	Reference
Blueberries:	Untargeted/	UHPLC-MS	Principal Component	Zhao et al.,
polyphenol	informative		Analysis (PCA)	2017
composition				
Cheese: Production	Untargeted/	IMS	Compound	Vautz et al.,
control	informative		identification	2006
Honey:	Untargeted/	NMR	Partial Least Square	Li et al.,
determinations of	discriminative/		(PLS)	2016
residues	predictive			
Meat: quality/safety	Targeted/	HILIC-MS/MS	MRL Validation	Dasenaki et
	discriminative			al., 2016
Olive oil:	Untargeted/	GC-HRMS	Orthogonal partial	Sales et al.,
classification	predictive		least squares	2017
			discriminant analysis	
			(OPLS-DA)	
Maize: GMO	Untargeted/	CE-TOF-MS	PCA	Levandi et
identification	discriminative			al., 2008
Vinegar:	Untargeted/	GC-MS	Hierarchical cluster	Pinu et al.,
Metabolic profile	discriminative		analysis (HCA)	2016
Fermented vegetable	Untargerted/	NMR	PCA	Tomita et
juice: monitoring	discriminative/		OPLS-DA	al., 2017
metabolites	predictive			
Coffee: determination	Untargeted/	UHPLC-(Q)ToF	PCA	Ossa et al.,
of product origin	predictive/	MS	PLS-DA	2018
	discriminative			
Almonds:	Untargerted/	UHPLC-(Q)ToF	PCA	Solsona et
discrimination	discriminative/		PLS-DA	al., 2018
	predictive			
Tea	Targeted	GC-MS	MRL Validation	Saito-Shida
				et al., 2015
Shellfish:	Targeted	LC/GC-MS-MS	Compound	Chang et
drug residue			identification	al., 2016
Wheat	Untargeted	UHPLC-(Q)ToF	PCA	Rubert et
			OPLS-DA	al., 2017a

Table 1. The metabolomics studies of different food samples

Metabolomics in Food Quality

The main target of food quality states is to ensure the appropriate organoleptic and desired characteristics of the product. The research related to food quality aims to evaluate the impacts of food processes on food compounds. The improvement in metabolomic techniques has allowed the monitoring of some quality parameters of foods during processing. The metabolomics methods have been commonly employed to identify the chemical composition of food products (Cossignani et al., 2014). The taste quality of white tea was evaluated by non-targeted metabolomics in combination with multivariate analysis (Yue et al., 2019). A metabolomic data-based prediction model was constructed which could separate green, yellow, and white teas according to unique tea metabolites (Zhang et al., 2019). The two-dimensional liquid chromatography was utilized in the determination of anthocyanins of different aged red wines and also to make the comparison of metabolites of aged red wines (Willemse et al., 2015). An untargeted metabolomics technique was used to distinguish the volatile compounds of grape juices according to various fungal infections performed by *Botrytis cinerea*, *Penicillium expansum*, *Aspergillus niger*, or *A*. carbonarius (Schueuermann et al., 2019).

Food authenticity and quality are closely related to each other. Food adulteration exposes defects in quality by incorporating prohibited ingredients or poor products, causing a health threat to customers. Therefore, food authenticity has great importance in the food industry for maintaining nutritional benefits, origin, and production standards. The metabolomics-based methods effectively specify the discrimination potential between adulterated and authentic foods. Moreover, metabolomics has been used to define the protected designation of origin (PDO) of food products. A prediction model was performed to discriminate adulterated Bordeaux wine in terms of geographical origin with LC-QTOF-MS (Lin et al., 2014). The different ingredients of the milk powders were analyzed by metabolomics coupled with 2DLC and monosaccharides were found in various trademarks and types of milk powder (Ma et al., 2014).

Metabolomics-based methods have been extensively used in examining the ingredients stated on the label of fruits, rice, and different vegetables (potatoes, peppers, white cabbage), clarifying differences in established production systems of wheat and maize, detecting the origin of olive oil, vinegar, wine, almonds, cocoa beans, honey, coffee species, saffron, and citrus fruit juices (Li et al., 2021).

Metabolomics in Food Microbiology

particularly Salmonella The foodborne pathogens, Listeria spp., Campylobacter jejuni, and Shigella spp., monocytogenes, spoilage microorganisms (such as Pseudomonas spp., Acinetobacter spp., and Botrytis spp.), and currently emerging pathogens have appeared as a result of the internationalization of the food supply chain and food consumption trends (Li and Zhu, 2017). Traditional laboratory techniques are primarily timeconsuming and labor-intensive. A large number of microbial biomarkers with varying levels of microbial contamination have been identified using metabolomics-based methods. These methods have demonstrated significant promise in the early detection of microbial contamination. The GC-MS has been used to characterize three prominent foodborne pathogens, E. coli O157:H7, L. monocytogenes, and S. enterica. Several potential pathogenspecific biomarkers were discovered, allowing the rapid differentiation of food samples containing pathogens (Jadhav et al., 2019). Furthermore, metabolomics has the ability to determine some antimicrobial compounds responsible for the antimicrobial characteristics of foods (Cevallos-Cevallos et al., 2009).

Despite the fact that this technique is promising in identifying metabolic shifts in different stages of microorganisms and providing important information on pathogen microorganisms, it is challenging to develop a universal method for removing bacterial cultures during metabolomics research. The extracellular and intracellular metabolites of microorganisms are typically difficult to distinguish from each other and also to identify. Moreover, metabolites are relatively unstable and could be quickly degraded during the handling and processing of samples. The current improvements in microbial metabolomics are not optimal. Therefore, more sensitive and reproducible analytical tools, as well as appropriate sample treatment techniques, are required to successfully determine the complexity of metabolites for metabolomics analysis (Li et al., 2021).Metabolomics in Food Safety

The foods could be contaminated with foodborne causative agents (pathogens and biotoxins), and some physical and chemical materials (pesticides and metals) which cause to foodborne illnesses throughout the critical steps of food processes. Metabolomics is able to evaluate the safety conditions of foods during processing and pre- and post-harvest periods and storage. Generally, untargeted discriminative analysis has been used for food safety. The MS/NMR-based metabolomics analysis is superior for the

245

detection and quantification of pathogens, chemical contaminants, prohibition compounds, and natural toxins (Rešetar et al., 2015).

Foodborne pathogens and their toxins do not remarkably the change characteristic properties (flavor, texture, or appearance) of foods. As a result, microbial contaminants are the most frequently reported foodborne causative agents. Microbial metabolomics may also be used to explain how environmental factors affect complex biological systems (Anderson et al., 2014). The metabolomic applications for food microbiology are discussed in the previous section.

Foodborne biotoxins are classified into two groups: intrinsic foodborne biotoxins and extrinsic foodborne biotoxins. These toxins are synthesized and released into the environment during pathogen growth and cause common foodborne outbreaks. Metabolomics is able to detect toxins in the early stages to ensure food safety. Mass spectroscopy, in particular, could profile metabolites associated with microbial contamination, and NMR could also directly define microbial toxins (Kleigrewe et al., 2012). Some natural toxins are known to be lethal since they are extremely toxic at lower concentrations. As a result, preparation methods should be improved to provide sensitive and high-throughput detection. For example, the addition of multiple antibody immunoaffinity columns prior to HPLC-MS/MS was selectively determined different toxins with a reduction in the detection limit (Zhang et al., 2016).

Fungicides, pesticides, antibiotics, and nanomaterials are examples of xenobiotics that can contaminate food via the atmosphere, soil, and water. These xenobiotics negatively affect whole living organisms even at low levels. Metabolomics can detect various groups of xenobiotics in various food matrices (Li et al., 2021).

The metabolomics methods have been developed to control veterinary drug residues and antibiotics in animals following alterations in metabolites in biological tissues (Kaufmann et al., 2015). Liquid chromatography combined with electrospray ionization and tandem mass spectrometry (LC-ESI-MS/MS) has been utilized in the quantification and identification of 115 different veterinary drug and pharmaceutical residues in milk powder, egg, and fish tissue (Dasenaki and Thomaidis, 2015). A targeted analysis has been performed by investigating 20 different antibiotic residues in dairy products including powdered milk, commercial milk, and raw milk (Wang et al., 2017).

The pesticides are widely used in agricultural productions. Pesticides are receiving much more attention since the presence of residues in foods involves health risks. For multi-residue pesticide analyses in various foods, the combination of QuEChERS (Quick, Easy, Cheap, Effective, Rugged, and

Safe) with solid-phase extraction (SPE) or solid-phase micro-extraction (SPME) has been achieved. The QuEChERS approach was optimized for the determination of mycotoxins and pesticides in coffee. It was observed that 117 pesticides and 30 mycotoxins were present in raw coffee (González-Curbelo et al., 2015; Reichert et al., 2018).

The new genetic engineering developments and genetically engineered (GE) crops with genomic modifications have been produced and allowed for the cultivation of genetically modified (GM) crops. The unknown health threat of GM foods is a source of health concerns among consumers, and they have been restricted in many countries. It is critical to estimate the addition of genetically modified foods and/or food ingredients to the food supply and to evaluate risk parameters using metabolomics techniques. Untargeted metabolomics is routinely involved in the differentiation of conventional foods, transgenic foods, and genetically modified foods (Cevallos-Cevallos et al., 2009; Li et al., 2021). There have been comprehensive researches related to wheat (Shewry et al., 2007), soybean (García-Villalba et al., 2008), barley (Kogel, 2010) and potato (Shepherd et al., 2015) differentiate genetic modification for food safety.

Metabolomics in Food Processing

Food processing comprises the all over changes in physical and chemical structure that are created by components of foods that could be detected by metabolomics. Informative metabolomics is able to determine undesired effects pre-processing and during processing including degradation of compounds and nutrients, and synthesis of new compounds like toxins (Cevallos-Cevallos et al., 2009). The fermentation process of cheonggukjang has been monitored by informative and discriminative untargeted analysis by using nuclear magnetic resonance, and the final products were separated according to the fermentation time (Choi et al., 2007). Metabolomics could also be used to decide the suitability of raw materials for a certain process. While some potato varieties are favored for frying, others are used in the baking industry. The flow infusion electrospray-ionization mass spectrometry (FI-ESI-MS) and compound identification were aided by GC-MS to determine the differences between potato varieties (Beckmann et al., 2007).

Targeted and untargeted metabolomics have been widely used to measure the effects of process parameters on the metabolite composition of foods. There are several metabolomics studies related to different food processes; the

effects of storage conditions on red wine (Arapitsas et al., 2016), the influences of blending and heating on Tiger nut milk (Rubert et al., 2017b), carrot, tomato, broccoli (Lopez-Sanchez et al., 2015), and banana (Chen et al., 2020), the impacts of energy stages on mung beans sprout (Chen et al., 2019), and the consequences of heat processing on Brassica vegetables (Hennig et al., 2014).

Metabolomics for Nutrition

Foodomics is a discipline that focuses on food and nutrition to explain the relationship between ingredients of food with quality and safety, daily diet, health, and diseases via developed omics technologies and chemometrics, bioinformatics, and biostatistics to enhance consumer's beneficence, health, and knowledge. The main subjects of foodomics could be summarized into three groups;

- 1. Human health (food consumption related to human health)
- 2. Food resources (plant and animal origin of food)
- 3. Food Processing (Picone et al., 2019; Zotti et al., 2016).

Nowadays, foodomics is used to better understand the molecular connections between human health, nutrition, and food. In particular, foodomics has been used for developing novel nutraceutical and functional foods. The effects of bee pollen (BP) on inflammatory bowel disease were investigated utilizing metabolomics analysis by using ultra-performance liquid chromatography in tandem with quadrupole time of flight-mass spectrometry (UPLC-Q-TOF/MS). It was discovered that the BP extract's regulatory mechanism protects cellular metabolic pathways from DSS-induced Caco-2 cell metabolism disorders (Li et al., 2019). The metabolomic methods were used to assess the anti-proliferative potential of Passiflora mollissima seeds on HT-29 human colon cancer cells. The results confirm that foodomics enabled the identification of genes, comprising polyamine and glutathione metabolism, or the inactivation of NUPR1 transcription could be involved in fluctuations in intracellular ceramide concentrations (Ballesteros-Vivas et al., 2020).

One of the most important and challenging aspects of nutrition is the monitoring food consumption of consumers. It is a simple way to precisely control the health of consumers by monitoring the food that a person has

consumed. The food frequency questionnaires (FFQs) fill out by consumers revealed some findings on the positive or negative effects of certain foods or diets. It has also been observed that FFQ recall for retrospective diet studies is not good enough. Moreover, FFO does not provide direct information about plasma levels of circulating nutrients or micronutrients (Wishart, 2008). Food consumption monitoring combined with metabolomics provides detailed knowledge of blood, urine, or saliva from volunteers in diet intervention studies. Foods are complex molecules and have many more components and metabolites. Food-specific biomarkers could be found in either blood or urine. The presence of these biomarkers proves the consumption of a certain food or the amount of the compound increases significantly over a period of time. It is known that the substantial difference between urine and blood is the percentages of metabolites to non-metabolites. Generally, urine has a significantly higher proportion of non-metabolites than blood. In a study on biomarkers of onion consumption, 11 relatively high abundance quercitin metabolites were discovered in urine, whereas only 5 lower abundance quercitin metabolites were found in the blood (Mullen et al., 2004). Therefore, metabolomic studies could be a solution to the problems of food consumption questionnaires used in probable diet objection or food interference studies.

Conclusion

Metabolomics has achieved great progress, with exciting findings associated with food analysis through the development of analytical techniques. The application of metabolomics in food processing, food quality control, and food safety is satisfactory with appropriate statistical analyses. Furthermore, targeted metabolomics should be improved with the automation, fast, and precision of entire metabolite quantitation. The untargeted metabolomics should be refined to control analytical data quality and develop a more reliable standardization of protocols. Comprehensive attention should be paid to food microbiology and nutrition for future aspects.

References

Alonso, A., Marsal, S., & Julià, A. (2015). Analytical methods in untargeted metabolomics: state of the art in 2015. *Frontiers in bioengineering and biotechnology*, 3, 23. https://doi.org/10.3389/fbioe.2015.00023.

- Anderson, J., Bogart, N., Clarke, A., Nelson, L., Warren, B., & Jespersen, L. (2014). Food safety management in the global food supply chain. *Perspectives in Public Health*, 134(4), 181-181. https://doi.org/10.1177/1757913914538139.
- Arapitsas, P., Della Corte, A., Gika, H., Narduzzi, L., Mattivi, F., & Theodoridis, G. (2016). Studying the effect of storage conditions on the metabolite content of red wine using HILIC LC–MS based metabolomics. *Food chemistry*, 197, 1331-1340. https://doi.org/10.1016/j.foodchem.2015.09.084.
- Ballesteros-Vivas, D., Alvarez-Rivera, G., León, C., Morantes, S. J., Ibánez, E., Parada-Alfonso, F., Cifuentes, A., & Valdés, A. (2020). Foodomics evaluation of the antiproliferative potential of Passiflora mollissima seeds. *Food Research International*, 130, 108938. https://doi.org/10.1016/j.foodres.2019.108938.
- Beckmann, M., Enot, D. P., Overy, D. P., & Draper, J. (2007). Representation, comparison, and interpretation of metabolome fingerprint data for total composition analysis and quality trait investigation in potato cultivars. *Journal of Agricultural and Food Chemistry*, 55(9), 3444-3451. https://doi.org/10.1021/jf0701842.
- Bedair, M., & Sumner, L. W. (2008). Current and emerging mass-spectrometry technologies for metabolomics. *TrAC Trends in Analytical Chemistry*, 27(3), 238-250. https://doi.org/10.1016/j.trac.2008.01.006.
- Cevallos-Cevallos, J. M., Reyes-De-Corcuera, J. I., Etxeberria, E., Danyluk, M. D., & Rodrick, G. E. (2009). Metabolomic analysis in food science: a review. *Trends in Food Science & Technology*, 20(11-12), 557-566. https://doi.org/10.1016/j.tifs.2009. 07.002.
- Chaleckis, R., Meister, I., Zhang, P., & Wheelock, C. E. (2019). Challenges, progress and promises of metabolite annotation for LC–MS-based metabolomics. *Current opinion in biotechnology*, 55, 44-50. https://doi.org/10.1016/j.copbio.2018.07.010.
- Chang, G. R., Chen, H. S., & Lin, F. Y. (2016). Analysis of banned veterinary drugs and herbicide residues in shellfish by liquid chromatography-tandem mass spectrometry (LC/MS/MS) and gas chromatography-tandem mass spectrometry (GC/MS/MS). *Marine Pollution Bulletin*, 113(1-2), 579-584. https://doi.org/10.1016/j.marpolbul. 2016.08.080.
- Chen, L., Wu, J. E., Li, Z., Liu, Q., Zhao, X., & Yang, H. (2019). Metabolomic analysis of energy regulated germination and sprouting of organic mung bean (Vigna radiata) using NMR spectroscopy. *Food Chemistry*, 286, 87-97. https://doi.org/10.1016/j. foodchem.2019.01.183.
- Chen, L., Zhao, X., Wu, J. E., He, Y., & Yang, H. (2020). Metabolic analysis of salicylic acid-induced chilling tolerance of banana using NMR. *Food Research International*, 128, 108796. https://doi.org/10.1016/j.foodres.2019.108796.
- Choi, H. K., Yoon, J. H., Kim, Y. S., & Kwon, D. Y. (2007). Metabolomic profiling of Cheonggukjang during fermentation by 1H NMR spectrometry and principal components analysis. *Process Biochemistry*, 42(2), 263-266. https://doi.org/10. 1016/j.procbio.2006.07.014.
- Cook, P. W., & Nightingale, K. K. (2018). Use of omics methods for the advancement of food quality and food safety. *Animal Frontiers*, 8(4), 33-41. https://doi.org/10.1093/ af/vfy024.

- Cossignani, L., Urbani, E., Simonetti, M. S., Maurizi, A., Chiesi, C., & Blasi, F. (2014). Characterisation of secondary metabolites in saffron from central Italy (Cascia, Umbria). *Food chemistry*, 143, 446-451. https://doi.org/10.1016/j.foodchem.2013. 08.020.
- Cozzolino, D., Flood, L., Bellon, J., Gishen, M., & De Barros Lopes, M. (2006). Combining near infrared spectroscopy and multivariate analysis as a tool to differentiate different strains of *Saccharomyces cerevisiae*: a metabolomic study. *Yeast*, 23(14-15), 1089-1096. https://doi.org/10.1002/yea.1418.
- Dasenaki, M. E., & Thomaidis, N. S. (2015). Multi-residue determination of 115 veterinary drugs and pharmaceutical residues in milk powder, butter, fish tissue and eggs using liquid chromatography-tandem mass spectrometry. *Analytica Chimica Acta*, 880, 103-121. https://doi.org/10.1016/j.aca.2015.04.013.
- Dasenaki, M. E., Michali, C. S., & Thomaidis, N. S. (2016). Analysis of 76 veterinary pharmaceuticals from 13 classes including aminoglycosides in bovine muscle by hydrophilic interaction liquid chromatography–tandem mass spectrometry. *Journal of Chromatography A*, 1452, 67-80. https://doi.org/10.1016/j.chroma.2016.05.031.
- Fraga-Corral, M., Carpena, M., Garcia-Oliveira, P., Pereira, A. G., Prieto, M. A., & Simal-Gandara, J. (2022). Analytical metabolomics and applications in health, environmental and food science. *Critical Reviews in Analytical Chemistry*, 52(4), 712-734. https://doi.org/10.1080/10408347.2020.1823811.
- Gallo, M., & Ferranti, P. (2016). The evolution of analytical chemistry methods in foodomics. *Journal of Chromatography A*, 1428, 3-15. https://doi.org/10.1016/j. chroma.2015.09.007.
- García-Villalba, R., León, C., Dinelli, G., Segura-Carretero, A., Fernández-Gutiérrez, A., Garcia-Cañas, V., & Cifuentes, A. (2008). Comparative metabolomic study of transgenic versus conventional soybean using capillary electrophoresis–time-of-flight mass spectrometry. *Journal of Chromatography A*, *1195*(1-2), 164-173. https://doi.org/10.1016/j.chroma.2008.05.018.
- González-Curbelo, M. Á., Socas-Rodríguez, B., Herrera-Herrera, A. V., González-Sálamo, J., Hernández-Borges, J., & Rodríguez-Delgado, M. Á. (2015). Evolution and applications of the QuEChERS method. *TrAC Trends in Analytical Chemistry*, 71, 169-185. https://doi.org/10.1016/j.trac.2015.04.012.
- Green, R. D., Qureshi, M. A., Long, J. A., Burfening, P. J., & Hamernik, D. L. (2007). Identifying the future needs for long-term USDA efforts in agricultural animal genomics. *International journal of biological sciences*, 3(3), 185. https://doi.org/10. 7150/ijbs.3.185.
- Hall, R. D., Brouwer, I. D., & Fitzgerald, M. A. (2008). Plant metabolomics and its potential application for human nutrition. *Physiologia plantarum*, 132(2), 162-175. https://doi.org/10.1111/j.1399-3054.2007.00989.x.
- Hennig, K., De Vos, R. C. H., Maliepaard, C., Dekker, M., Verkerk, R., & Bonnema, G. (2014). A metabolomics approach to identify factors influencing glucosinolate thermal degradation rates in Brassica vegetables. *Food chemistry*, 155, 287-297. https://doi.org/10.1016/j.foodchem.2014.01.062.
- Jadhav, S. R., Shah, R. M., Karpe, A. V., Beale, D. J., Kouremenos, K. A., & Palombo, E. A. (2019). Identification of putative biomarkers specific to foodborne pathogens using

251

metabolomics. In *Foodborne bacterial pathogens* (pp. 149-164). Humana, New York, NY.

- Kaufmann, A., Butcher, P., Maden, K., Walker, S., & Widmer, M. (2015). Reliability of veterinary drug residue confirmation: high resolution mass spectrometry versus tandem mass spectrometry. *Analytica Chimica Acta*, 856, 54-67. https://doi.org/10. 1016/j.aca.2014.11.034.
- Kleigrewe, K., Aydin, F., Hogrefe, K., Piecuch, P., Bergander, K., Würthwein, E. U., & Humpf, H. U. (2012). Structure elucidation of new fusarins revealing insights in the rearrangement mechanisms of the Fusarium mycotoxin fusarin C. *Journal of agricultural and food chemistry*, 60(21), 5497-5505. https://doi.org/10.1021/jf 3009469.
- Kogel, K. H. (2010). Transcriptome and metabolome profiling of field-grown transgenic barley lack induced differences but show cultivar-specific variances. *Proceedings of the National Academy of Sciences*, 107(14), 6198-6203. https://doi.org/10.1073/ pnas.1001945107.
- Levandi, T., Leon, C., Kaljurand, M., Garcia-Canas, V., & Cifuentes, A. (2008). Capillary electrophoresis time-of-flight mass spectrometry for comparative metabolomics of transgenic versus conventional maize. *Analytical chemistry*, 80(16), 6329-6335. https://doi.org/10.1021/ac8006329.
- Li, H., & Zhu, J. (2017). Targeted metabolic profiling rapidly differentiates Escherichia coli and Staphylococcus aureus at species and strain level. *Rapid Communications in Mass Spectrometry*, 31(19), 1669-1676. https://doi.org/10.1002/rcm.7949.
- Li, Q., Liang, X., Guo, N., Hu, L., Prasad, M. P., Wu, Y., Xue, X., Wu, L., & Wang, K. (2019). Protective effects of Bee pollen extract on the Caco-2 intestinal barrier dysfunctions induced by dextran sulfate sodium. *Biomedicine & Pharmacotherapy*, 117, 109200. https://doi.org/10.1016/j.biopha.2019.109200.
- Li, S., Tian, Y., Jiang, P., Lin, Y., Liu, X., & Yang, H. (2021). Recent advances in the application of metabolomics for food safety control and food quality analyses. *Critical Reviews in Food Science and Nutrition*, 61(9), 1448-1469. https://doi.org/10.1080/10408398.2020.1761287.
- Li, Y., Zhang, J., Jin, Y., Wang, L., Zhao, W., Zhang, W., Zhai, L., Zhang, Y., Zhang, Y., & Zhou, J. (2016). Hybrid quadrupole-orbitrap mass spectrometry analysis with accurate-mass database and parallel reaction monitoring for high-throughput screening and quantification of multi-xenobiotics in honey. *Journal of Chromatography A*, 1429, 119-126. https://doi.org/10.1016/j.chroma.2015.11.075.
- Lin, L., Xu, D., Shen, X., Zhang, Z., Chen, L., & Wang, C. (2014). Traceability of the wine in Bordeaux region from France based on liquid chromatography-quadrupole-time of flight tandem mass spectrometry. *Journal of Food Safety and Quality*, 5(9), 2649-2656.
- Lopez-Sanchez, P., De Vos, R. C. H., Jonker, H. H., Mumm, R., Hall, R. D., Bialek, L., Leenman, R., Strassburg, K., Vreeken, R., Hankemeier, T., Schumm, S., & Van Duynhoven, J. (2015). Comprehensive metabolomics to evaluate the impact of industrial processing on the phytochemical composition of vegetable purees. *Food Chemistry*, 168, 348-355. https://doi.org/10.1016/j.foodchem.2014.07.076.

- Ma, J., Hou, X., Zhang, B., Wang, Y., & He, L. (2014). The analysis of carbohydrates in milk powder by a new "heart-cutting" two-dimensional liquid chromatography method. *Journal of Pharmaceutical and Biomedical Analysis*, 91, 24-31. https://doi. org/10.1016/j.jpba.2013.11.006.
- Mullen, W., Boitier, A., Stewart, A. J., & Crozier, A. (2004). Flavonoid metabolites in human plasma and urine after the consumption of red onions: analysis by liquid chromatography with photodiode array and full scan tandem mass spectrometric detection. *Journal of Chromatography A*, 1058(1-2), 163-168. https://doi.org/10. 1016/j.chroma.2004.08.117.
- Ossa, D. E. H., Gil-Solsona, R., Peñuela, G. A., Sancho, J. V., & Hernández, F. J. (2018). Assessment of protected designation of origin for Colombian coffees based on HRMSbased metabolomics. *Food chemistry*, 250, 89-97. https://doi.org/10.1016/j. foodchem.2018.01.038.
- Picone, G., De Noni, I., Ferranti, P., Nicolai, M. A., Alamprese, C., Trimigno, A., Brodkorb, A., Portmann, R., El, S. H., & Capozzi, F. (2019). Monitoring molecular composition and digestibility of ripened bresaola through a combined foodomics approach. *Food Research International*, 115, 360-368. https://doi.org/10.1016/j. foodres.2018.11.021.
- Pinu, F. R. (2015). Metabolomics The new frontier in food safety and quality research. Food Research International, 72, 80-81. https://doi.org/10.1016/j.foodres.2015.03 .028.
- Pinu, F. R., de Carvalho-Silva, S., Trovatti Uetanabaro, A. P., & Villas-Boas, S. G. (2016). Vinegar metabolomics: An explorative study of commercial balsamic vinegars using gas chromatography-mass spectrometry. *Metabolites*, 6(3), 22. https://doi.org/10.3390/metabo6030022.
- Reichert, B., de Kok, A., Pizzutti, I. R., Scholten, J., Cardoso, C. D., & Spanjer, M. (2018). Simultaneous determination of 117 pesticides and 30 mycotoxins in raw coffee, without clean-up, by LC-ESI-MS/MS analysis. *Analytica Chimica Acta*, 1004, 40-50. https://doi.org/10.1016/j.aca.2017.11.077.
- Rešetar, D., Pavelić, S. K., & Josić, D. (2015). Foodomics for investigations of food toxins. *Current Opinion in Food Science*, 4, 86-91. https://doi.org/10.1016/j.cofs. 2015.05.004.
- Rubert, J., Monforte, A., Hurkova, K., Pérez-Martínez, G., Blesa, J., Navarro, J. L., Stranka, M., Soriano, J. M., & Hajslova, J. (2017b). Untargeted metabolomics of fresh and heat treatment Tiger nut (Cyperus esculentus L.) milks reveals further insight into food quality and nutrition. *Journal of chromatography A*, 1514, 80-87. https://doi.org/10.1016/j.chroma.2017.07.071.
- Rubert, J., Righetti, L., Stranska-Zachariasova, M., Dzuman, Z., Chrpova, J., Dall'Asta, C., & Hajslova, J. (2017a). Untargeted metabolomics based on ultra-high-performance liquid chromatography-high-resolution mass spectrometry merged with chemometrics: A new predictable tool for an early detection of mycotoxins. *Food chemistry*, 224, 423-431. https://doi.org/10.1016/j.foodchem.2016.11.132.
- Saito-Shida, S., Nemoto, S., & Teshima, R. (2015). Multiresidue determination of pesticides in tea by gas chromatography-tandem mass spectrometry. *Journal of*

Environmental Science and Health, Part B, 50(11), 760-776. https://doi.org/10.1080/03601234.2015.1058092.

- Sales, C., Cervera, M. I., Gil, R., Portolés, T., Pitarch, E., & Beltran, J. (2017). Quality classification of Spanish olive oils by untargeted gas chromatography coupled to hybrid quadrupole-time of flight mass spectrometry with atmospheric pressure chemical ionization and metabolomics-based statistical approach. *Food chemistry*, 216, 365-373. https://doi.org/10.1016/j.foodchem.2016.08.033.
- Schueuermann, C., Steel, C. C., Blackman, J. W., Clark, A. C., Schwarz, L. J., Moraga, J., Collado, I. G., & Schmidtke, L. M. (2019). A GC–MS untargeted metabolomics approach for the classification of chemical differences in grape juices based on fungal pathogen. *Food chemistry*, 270, 375-384. https://doi.org/10.1016/j.foodchem. 2018.07.057.
- Shepherd, L. V. T., Hackett, C. A., Alexander, C. J., McNicol, J. W., Sungurtas, J. A., Stewart, D., McCue, K. F., Belknap, W. R., & Davies, H. V. (2015). Modifying glycoalkaloid content in transgenic potato–Metabolome impacts. *Food chemistry*, 187, 437-443. https://doi.org/10.1016/j.foodchem.2015.04.111.
- Shewry, P. R., Baudo, M., Lovegrove, A., Powers, S., Napier, J. A., Ward, J. L., Baker, J. M., & Beale, M. H. (2007). Are GM and conventionally bred cereals really different? *Trends in Food Science & Technology*, 18(4), 201-209. https://doi.org/10. 1016/j.tifs.2006.12.010.
- Solsona, R., Boix, C., Ibáñez, M., & Sancho, J. V. (2018). The classification of almonds (Prunus dulcis) by country and variety using UHPLC-HRMS-based untargeted metabolomics. *Food Additives & Contaminants: Part A*, 35(3), 395-403. https://doi. org/10.1080/19440049.2017.1416679.
- Tomita, S., Saito, K., Nakamura, T., Sekiyama, Y., & Kikuchi, J. (2017). Rapid discrimination of strain-dependent fermentation characteristics among Lactobacillus strains by NMR-based metabolomics of fermented vegetable juice. *PLoS One*, 12(7), e0182229. https://doi.org/10.1371/journal.pone.0182229.
- Vautz, W., Zimmermann, D., Hartmann, M., Baumbach, J. I., Nolte, J., & Jung, J. (2006). Ion mobility spectrometry for food quality and safety. *Food additives and contaminants*, 23(11), 1064-1073. https://doi.org/10.1080/02652030600889590.
- Wang, L., Yang, B., Zhang, X., & Zheng, H. (2017). Novel two-dimensional liquid chromatography-tandem mass spectrometry for the analysis of twenty antibiotics residues in dairy products. *Food Analytical Methods*, 10(6), 2001-2010. 1–2010 https://doi.org/10.1007/s12161-016-0763-4.
- Willemse, C. M., Stander, M. A., Vestner, J., Tredoux, A. G., & De Villiers, A. (2015). Comprehensive two-dimensional hydrophilic interaction chromatography (HILIC)× reversed-phase liquid chromatography coupled to high-resolution mass spectrometry (RP-LC-UV-MS) analysis of anthocyanins and derived pigments in red wine. *Analytical chemistry*, 87(24), 12006-12015. https://doi.org/10.1021/acs.analchem. 5b03615.
- Wishart, D. S. (2008). Metabolomics: applications to food science and nutrition research. *Trends in food science & technology*, 19(9), 482-493. https://doi.org/10.1016/j.tifs. 2008.03.003.

- Yue, W., Sun, W., Rao, R. S. P., Ye, N., Yang, Z., & Chen, M. (2019). Non-targeted metabolomics reveals distinct chemical compositions among different grades of Bai Mudan white tea. *Food chemistry*, 277, 289-297. https://doi.org/10.1016/j.foodchem. 2018.10.113.
- Zhang, A., Sun, H., Wang, P., Han, Y., & Wang, X. (2012). Modern analytical techniques in metabolomics analysis. *Analyst*, 137(2), 293-300. https://doi.org/10.1039/C1AN 15605E.
- Zhang, Q., Wu, S., Li, Y., Liu, M., Ni, K., Yi, X., Shi, Y., Ma, L., Willmitzer, L., & Ruan, J. (2019). Characterization of three different classes of non-fermented teas using untargeted metabolomics. *Food Research International*, 121, 697-704. https://doi.org/10.1016/j.foodres.2018.12.042.
- Zhang, Z., Hu, X., Zhang, Q., & Li, P. (2016). Determination for multiple mycotoxins in agricultural products using HPLC–MS/MS via a multiple antibody immunoaffinity column. *Journal of Chromatography B*, 1021, 145-152. https://doi.org/10.1016/j. jchromb.2016.02.035.
- Zhao, Y., Wu, X., Yu, L., & Chen, P. (2017). Retention of polyphenols in blueberries (Vaccinium corymbosum) after different cooking methods, using UHPLC–DAD–MS based metabolomics. *Journal of Food Composition and Analysis*, 56, 55-66. https://doi.org/10.1016/j.jfca.2016.12.003.
- Zotti, M., Del Coco, L., De Pascali, S. A., Migoni, D., Vizzini, S., Mancinelli, G., & Fanizzi, F. P. (2016). Comparative analysis of the proximate and elemental composition of the blue crab Callinectes sapidus, the warty crab Eriphia verrucosa, and the edible crab Cancer pagurus. *Heliyon*, 2(2), e00075. https://doi.org/10. 1016/j.heliyon.2016.e00075.

255